

Mefepronic acid, a PPAR agonist, is inefficient on reproductive performance of ewes in both early and late postpartum period



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SUMMARY

The liver is a dynamic organ that plays critical roles in many physiological processes. In farm animals, supporting the liver reduces postpartum metabolic diseases and loss of reproductive and milk yield in the transition period. The aim of this study is to investigate the effects of mefepronic acid (2-methyl-2-phenoxy propionic acid) injection on reproductive performance parameters with short-term progesterone administration in early and late postpartum period Hungarian Merino ewes during the non-breeding season. In the study, 89 adult ewes in the early (n=45) and late (n=44) postpartum (pp) period were used in the study. Control and treatment (MA) groups were formed separately for each pp period. Following insertion of a vaginal sponge containing 60 mg medroxyprogesterone acetate for 7 days, PMSG 500 IU was injected intramuscularly (day 7) to all ewes. In treatment groups (Early MA and Late MA), 10 mg/kg mefepronic acid (Hepagen®, Fatro Günesli) was injected (day 7), and remaining ewes consisted of control groups (Early Control and Late Control). The estrus signs of the ewes were followed and they were hand-mated (ewe: ram=5:1). Pregnancies were determined with transabdominal real time B-Mode ultrasonography with convex probe (3.5 MHz) on day 45 post-mating. There were no statistical differences in estrus rate, pregnancy rate, lambing rate and litter size at 1st service, 2nd service and overall services in early and late pp groups. According to results of this study, administration of mefepronic acid to ewes in the early and late pp period had no increasing effect on reproductive parameters. However, further studies investigating the survival of embryos and prostaglandin metabolism are needed to determine the efficacy of mefepronic acid in ewes.

KEY WORDS

Ewes; Mefepronic acid; Progesterone; Postpartum; Reproductive parameters.

INTRODUCTION

The two most popular accelerated lambing programs are the “three lambings in two years”¹ and the Cornell STAR® system (five lambings in three years)². If these programs are implemented, lambing is possible every 8 months or 7.2 months, respectively. For success in breeding, it is essential to keep exposures brief to reduce the time between lambing, and lambs should be weaned at two months old to enable breeding to occur again³. The act of boosting the intake of nutrients and improving overall condition prior to and during breeding is known as «flushing. Its aim is to raise the rate of ovulation, resulting in an increased lambing rate. Flushing is especially advantageous for skinny ewes that have not rebounded from the stress of previous lactation⁴. The delay in weaning of the ewes, which in practice takes 60-90 days, and when it is time to mate immediately, a dilemma will arise. On the one hand, it is necessary not to give the feed for weaning, on the other hand, it is necessary

to increase the feed before mating to increase the ovulation rate. During this period, there may be a possibility of body fat mobilization⁵, and the liver cannot fully metabolize them⁶. At present, there exist alternative approaches aimed at diminishing the incidence of metabolic disorders⁷. The intention behind these alternative options is to minimize the removal of body fat and the buildup of triacylglycerols in liver tissue, in order to preserve its structure, function, and prevent disturbances in the energy metabolism⁸. Several genes that regulate lipid metabolism and adipocyte differentiation are controlled by the activation of the peroxisome proliferator-activated receptors (PPARs), which belong to the superfamily of nuclear receptors⁹. PPARs are categorized into three isotypes: α , β , and γ . PPARs play a role in regulating lipid metabolism¹⁰⁻¹³. PPAR is triggered by fatty acids, prostaglandins, and fibrates, which are drugs utilized in human medicine for their ability to lower lipid levels¹⁴. Activation of PPAR promotes mitochondrial activation, peroxisomal β -oxidation, hepatic gluconeogenesis, metabolism of lipoproteins, choleric and cholagogic activity¹⁵. PPAR interferes with different steps of the inflammatory response such as cytokines (ILs, TNF, IFN γ), cytokine receptors, adhesion molecules, acute phase proteins (fibrinogen, C-reactive protein)¹⁵. Many bovine studies have found that they lower levels of NEFA,

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BHB, AST and other biochemical parameters in transition period^{16,17}. There is a close relationship between these hepatic metabolism products and reproduction. In addition, metabolomics researches have shown that lack of certain nutrients directly affect metabolic diseases and thus reproduction in cows and ewes^{18,19}.

Mefepronic acid (2-methyl-2-phenoxy propionic acid) belongs to the fibrate family of substances that are utilized in treating dyslipidemia²⁰, which is used in veterinary medicine¹⁵. Mefepronic acid promotes the liver's physiological functions and activates digestive processes²¹. Although the effects of mefepronic acid on the reproduction of ewes are not known, it has been reported to have positive effects in limited studies on cows^{6,7}. Rizzo et al.⁶ have emphasized the beneficial effects of mefepronic acid on the hepatic metabolism and reproductive parameters of postpartum dairy cows.

It was thought that the use of mefepronic acid could prevent the negative effects of not feeding the ewes for weaning and reduce lipomobilization in skinny ewes in the early postpartum period. In order to increase fertility within the framework of the mentioned above, the present study was aimed to investigate the effects of mefepronic acid on reproductive performance parameters with short-term progesterone administration in early and late postpartum period Hungarian Merino ewes during the non-breeding season.

MATERIAL AND METHODS

The present study was conducted with approval from Çukurova University Animal Experiments Local Ethics Committee, Adana, Türkiye (2022, 5/4).

Animals

This study was conducted in a commercial sheep farm (Lat: 37° 43' 94.80» N, Long: 35° 73' 37.90» E and Alt: 120 m) in Adana province in Türkiye during the non-breeding season (June) in 2022. A total of 89 adult Hungarian Merino ewes, clinically healthy, 2-3 years old, having 55-75 kg body weight were used. Ewes were grazed on artificial pasture 12 h of a day and not given any compound feed. Water was given ad libitum. No nutritional flushing was applied to animals before mating. Lambs were completely weaned 7 days before estrus stimulation.

Study design

The study design is illustrated in Figure 1. Ewes were grouped as Early Control (Day pp 35-60; 50.6 d ± 1.8; n=22), Late Control (Day pp 60-112; 85.2 d ± 3.4; n=22), Early MA (Day pp 37-59; 51.2 d ± 1.7; n=23) and Late MA (Day pp 60-117; 89.2 d ± 3.2; n=22) in the study.

Following insertion of vaginal sponge containing 60 mg medroxyprogesterone acetate (Esponjavet®, Hipra, Spain) for 7 day, PMSG 500 IU (Oviser®, Hipra) was injected intramuscularly (IM) (day 7) to all ewes. In treatment groups (Early MA and Late MA), 10 mg/kg mefepronic acid (100 mg/mL, 2-methyl-2-phenoxy propionic acid, Hepagen®, Fatro Günesli) was injected intramuscularly (Day 7). One teaser ram was used twice a day for 1 h duration for 24 h after removal of sponges for estrus detection. Ewes determined to be in estrus were hand-mated with one of the proven fertile Merino rams (ewe: ram ratio of 5:1). In all ewes, transabdominal ultrasound examination (Hitachi EUB-405, 3.5 MHz convex probe) was performed to diagnose pregnancy on day 45 post-mating. The litter size was determined at parturition.

Estrus detection rate, pregnancy rate, lambing rate and litter size were calculated as reproductive parameters as follows;

Statistical analysis

All statistical analyses were performed by using the SAS Version 8.2 (2001). For analyzing the calculated reproductive parameters, Chi-squared test, Fisher's exact test and GLIMMIX procedure were used. Proportional data was analyzed with Chi-squared test, Fisher's exact test between in early and late postpartum period within the same service period. Estrus rate, pregnancy rate and lambing rate were analyzed with GLIMMIX procedure between 1st, 2nd and overall service periods. The results were given as the percentage or mean ± standard error of the mean (SEM). Calculated P values less than 0.05 were considered significant.

RESULTS

At 1st service, 2nd service and overall service, estrus rate, pregnancy rate, lambing rate, number of lambs and litter size were determined in Early Control, Late Control, Early MA and Late MA groups. There were no statistical differences (p>0.05) in all reproductive parameters among the groups. Reproductive

Table 1 - Reproductive parameters in Control and Mefepronic acid (MA) groups in early and late pp period at the end of the study.

	At 1 st service				P	At 2 nd Service				P	Overall serviced ewes				P
	Early Control Group (n=22)	Late Control Group (n=22)	Early MA Group (n=23)	Late MA Group (n=22)		Early Control Group (n=11)	Late Control Group (n=9)	Early MA Group (n=8)	Late MA Group (n=10)		Early Control Group (n=22)	Late Control Group (n=22)	Early MA Group (n=23)	Late MA Group (n=22)	
Estrus Rate	86.4% (19/22)	100% (22/22)	82.6% (19/23)	90.9% (20/22)	0.24	72.7% (8/11)	88.9% (8/9)	62.5% (5/8)	90.0% (9/10)	0.41	86.4% (19/22)	95.5% (21/22)	87.0% (20/23)	95.5% (21/22)	0.55
Pregnancy Rate	50.0% (11/22)	59.1% (13/22)	65.2% (15/23)	54.5% (12/22)	0.76	72.7% (8/11)	88.9% (8/9)	62.5% (5/8)	90.0% (9/10)	0.41	86.4% (19/22)	95.5% (21/22)	87.0% (20/23)	95.5% (21/22)	0.55
Lambing Rate	100% (11/11)	100% (13/13)	100% (15/15)	100% (12/12)	>0.99	100% (8/8)	100% (8/8)	100% (5/5)	100% (9/9)	>0.99	100% (19/19)	100% (21/21)	100% (20/20)	100% (21/21)	<0.99
Number of Kids	14	17	21	16		8	10	5	10		22	27	26	26	
Single	8	9	9	8		8	6	5	8		16	15	14	16	
Twin	3 (6)	4 (8)	6 (12)	4 (8)		-	2 (4)	-	1 (2)		3 (6)	6 (12)	6 (12)	5 (10)	
Litter Size	1.27 (14/11)	1.31 (17/13)	1.4 (21/15)	1.33 (16/12)	0.99	1.0 (8/8)	1.25 (10/8)	1.0 (5/5)	1.11 (10/9)	0.98	1.27 (22/19)	1.29 (27/21)	1.30 (26/20)	1.24 (26/21)	0.99

P>0.05 is considered as non significant.

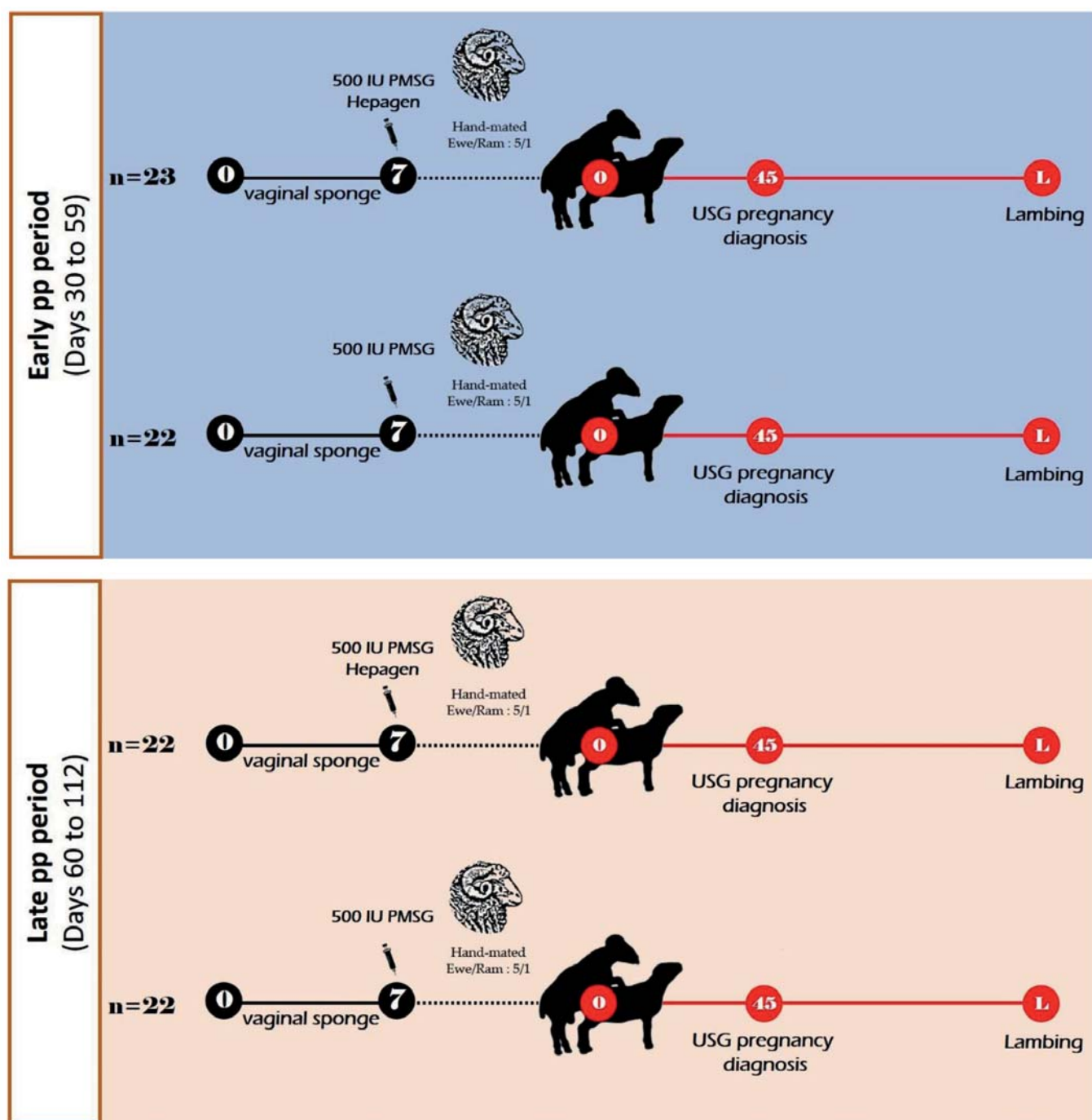


Figure 1 - The schematic illustration of the study. Blue and Pink diagrams represent the Early and Late postpartum period, respectively. Ewes with the same care and feeding conditions in the early and late postpartum periods were used in the study. Mefepronic acid was administered at a dose of 10 mg/kg (100 mg/mL, 2-methyl-2-phenoxy propionic acid, Hepagen®, Fatro Günesli) in the trial groups. Mefepronic acid injections were made on the day of PMSG administration.

parameters are presented in Table 1.

Although a numerical difference in pregnancy rates was determined between the Early Control group (50%) and the Early MA group (65.2%), this difference was not statistically significant.

DISCUSSION

Impairment of reproductive effectiveness is related to hepatic dysfunction (lipidosis and increasing hepatic clearance) through a complicated network of mutual interactions in dairy

cows²². In this context, supporting hepatic activities may help improve the reproductive performance in ewes. Mefepronic acid (MA), a PPAR agonist, capable of promoting peroxisomal β -oxidation and hepatic gluconeogenesis, was utilized in this study. In this current study, we have investigated, for the first time, the efficacy of mefepronic acid on reproductive parameters in ewes during early and late postpartum period.

It is well known that intravaginal sponges have been a choice of traditional treatments for estrus induction out of the breeding seasons in ewes²³. In short term (6-7 days) progestagen applications out of the breeding season, estrus rates were in ewes have been reported by Sareminejad et al.²⁴ (93.3%),

Özyurtlu et al.²⁵ (91.6%) and Yilmazer²⁶ (97%). We have previously reported the rate of estrus as 93.4% and 91.1% in Hungarian Merino ewes to which they applied sponges containing medroxyprogesterone acetate for 7 days with 500 IU PMSG²⁷. In this study, there were no statistical differences ($p>0.05$) at 1st service between the early control, late control, early MA and late MA groups in terms of estrus rate, 86.4%, 100%, 82.6% and 90.9%, respectively.

Pregnancy rate is one of the most important parameters in the evaluation of reproductive performance and economic profitability in farms. Rizzo et al.⁶ have reported that the calving to 1st estrus interval was shorter as it was recorded 74 ± 6.2 days in the control group and 50 ± 3 days in the MA group. Also pregnancy rate increased by 7% in the first and 7% second inseminations of MA applied Friesian cows compared to the control group. In this study, in which the efficacy of mefepronnic acid in ewes in the postpartum period was evaluated, it was found no statistical differences in pregnancy rate, litter size at 1st service among the groups (Table 1).

However, the pregnancy rate and litter size in the Early MA group was numerically higher at 1st service. These numerical differences could be attributed to two different mechanisms. First, mefepronnic acid increases the cholesterol level, which participates in the structure of progesterone. Steroid synthesis commonly begins with cholesterol, which can either be obtained from the diet or produced within the body and transported to the ovaries via lipoproteins (HDL and LDL)²⁸. The step that limits the rate of progesterone synthesis is the transportation of cholesterol into the mitochondrion²⁹. Progesterone plays a key role in regulation of the implantation of the blastocyst³⁰, and is necessary at a certain concentration after post-mating, as ewes with lower progesterone concentration suffer greater embryo loss³¹. Rizzo et al.⁶ showed that mefepronnic acid administration increases cholesterol and HDL levels and thus supports hepatic metabolism and lipogenesis, with an inverse trend to NEFA. The second is with the COX-2 mechanism. Some researchers³²⁻³⁴ have shown that PPAR can inhibit COX-2 expression. PPAR agonists are able to inhibit the induction of interleukin-6 (IL-6) and cyclooxygenase-2 (COX-2) by interleukin-1 (IL-1)³². COX-2 is also involved in the production of prostaglandin F₂ alpha. Prostaglandin F₂ alpha is the most important factor that controls regression of the corpus luteum³⁵. With mefepronnic acid injection, it is aimed to prolong the life of the corpus luteum (CL) by suppressing the release of PGF₂α during the maternal recognition period. Perhaps, by suppressing the release of PGF₂α during the maternal recognition period with mefepronnic acid injection, the lifespan of the corpus luteum (CL) can be extended. Rizzo et al.⁶ have reported that it increased the progesterone concentration in cows treated with mefepronnic acid on the 13th day after insemination compared to the control group (6.47 ± 0.37 vs. 4.24 ± 0.37 ng/mL). In parallel, mefepronnic acid has been shown to modulate PPAR expression in bovine liver based on histological results⁶.

Although it has been shown that there is an improvement in reproductive parameters with the application of mefepronnic acid in the postpartum period in cows^{6,7,15}, no clinical efficacy of mefepronnic acid has been observed in ewes in the early and late postpartum period out of the breeding season. This may be due to the fact that the hepatic metabolism in sheep is not as intense as in cows. For this reason, we suggest that the study should be repeated in sheep herds with high lactation efficiency and intensive feeding conditions.

CONCLUSION

In conclusion, administration of mefepronnic acid to ewes in the early and late postpartum period had no increasing effect on reproductive parameters. However, we think that further studies investigating the embryo survival and prostaglandin metabolism are needed to determine the efficacy of mefepronnic acid in ewes, especially with high lactation efficiency and intensive feeding conditions.

Author contribution

Conceptualization: Metehan Kutlu; data curation: Metehan Kutlu; formal analysis and investigation: Metehan Kutlu; funding acquisition: Metehan Kutlu; investigation: Metehan Kutlu; methodology: Metehan Kutlu; project administration: Metehan Kutlu; resources: Metehan Kutlu; software: Halef Dogan; supervision: Metehan Kutlu, Halef Dogan; validation: Metehan Kutlu, Halef Dogan; visualization: Halef Dogan; writing-original draft preparation: Metehan Kutlu, Halef Dogan, Eray Aktug; writing-review and editing: Metehan Kutlu, Halef Dogan, Eray Aktug.

Data availability

Not applicable.

Code availability

Not applicable.

Conflict of interest

The authors declare no competing interests.

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References

1. Alaçam, E. 1993. Koyunlarda siklik düzen ve üremenin denetlenmesi. Hayvancılık Ara. Derg., 3:65-69.
2. Smith, M.C. 2006. Veterinary experiences with the Cornell STAR system of accelerated lambing. Small Ruminant Research, 62:125-128.
3. Lewis, R.M., Notter, D.R., Hogue, D.E., Magee, B.H. 1996. Ewe fertility in the STAR accelerated lambing system. J. Anim. Sci., 74:1511-1522.
4. Shad, F.I., Tufani, N.A., Ganie, A.M., Ahmed, H.A. 2011. Flushing in Ewes for Higher Fecundity and Fertility. Livestock International, 15(2):10-14.
5. Raoofi, A., Jafarian, M., Safi, S., Vatankhah, M. 2013. Fluctuations in energy-related metabolites during the peri-parturition period in Lori-Bakhtiari ewes. Small Ruminant Research, 109:64-68.
6. Rizzo, A., Gazza, C., Mutinati, M., Desantis, S., Zizza, S., D'Onghia, G., D'Onghia, G., Pantaleo, M., Sciorsci, R.L. 2014. Effects of mefepronnic acid (2-phenoxy-2-methyl propionic acid) on hepatic metabolism and reproductive parameters in postpartum dairy cows. Endocr. Metab. Immune Disord. Drug Targets, 14:113-122.
7. Aparicio-Cecilio, A., Bouda, J., Salgado-Hernández, E.G., Núñez-Ochoa, L., Castillo-Mata, D.A., Gutiérrez-Chávez, A.J. 2012. Effect of 2-methyl-2-phenoxy propionic acid on serum lipid profile and ovarian activity in dairy cows. Czech Journal of Animal Science, 57: 550-556.
8. Grummer, R.R. 2008. Nutritional and management strategies for the prevention of fatty liver in dairy cattle. Vet. J., 176:10-20.
9. Willson, T.M., Brown, P.J., Sternbach, D.D., Henke, B.R. 2000. The PPARs: From Orphan Receptors to Drug Discovery. Journal of Medicinal Chemistry, 43:527-550.
10. Ahmed, W., Ziouzenkova, O., Brown, J., Devchand, P., Francis, S.,

- Kadokia, M., Kanda, T., Orasanu, G., Sharlach, M., Zandbergen, F., Plutzky, J. 2007. PPARs and their metabolic modulation: new mechanisms for transcriptional regulation?. *J. Intern. Med.*, 262:184-198.
11. Bionaz, M., Baumrucker, C.R., Shirk, E., Vanden Heuvel, J.P., Block, E., Varga, G.A. 2008. Characterization of Madin-Darby bovine kidney cell line for peroxisome proliferator-activated receptors: temporal response and sensitivity to fatty acids. *J. Dairy Sci.*, 91:2808-2813.
 12. Inoue, H., Jiang, X.F., Katayama, T., Osada, S., Umesono, K., Namura, S. 2003. Brain protection by resveratrol and fenofibrate against stroke requires peroxisome proliferator-activated receptor alpha in mice. *Neurosci. Lett.*, 352:203-206.
 13. Kota, B.P., Huang, T.H., Roufogalis, B.D. 2005. An overview on biological mechanisms of PPARs. *Pharmacol. Res.*, 51:85-94.
 14. Yoshikawa, T., Ide, T., Shimano, H., Yahagi, N., Amemiya-Kudo, M., Matsuzaka, T., Yatoh, S., Kitamine, T., Okazaki, H., Tamura, Y., Sekiya, M., Takahashi, A., Hasty, A.H., Sato, R., Sone, H., Osuga, J., Ishibashi, S., Yamada, N. 2003. Cross-talk between peroxisome proliferator-activated receptor (PPAR) alpha and liver X receptor (LXR) in nutritional regulation of fatty acid metabolism. I. PPARs suppress sterol regulatory element binding protein-1c promoter through inhibition of LXR signaling. *Mol. Endocrinol.*, 17:1240-1254.
 15. Sciorsci, R.L. 2018. Clinical approach to metabolic and reproductive pathologies: 1. In vivo and in vitro activity of mifepronic acid in postpartum dairy cows. Pages 329-330 in Proceedings of the National, 5th Herd Health & Management Congress, Antalya, Turkey.
 16. Fiore, E., Perillo, L., Giancesella, M., Giannetto, C., Giudice, E., Piccione, G., Morgante, M. 2021. Comparison between two preventive treatments for hyperketonaemia carried out pre-partum: effects on non-esterified fatty acids, -hydroxybutyrate and some biochemical parameters during peripartum and early lactation. *Journal of Dairy Research*, 88(1):38-44.
 17. Fiore, E., Perillo, L., Piccione, G., Giancesella, M., Bedin, S., Armato, L., Giudice, E., Morgante, M. 2016. Effect of combined acetylmethionine, cyanocobalamin and -lipoic acid on hepatic metabolism in high-yielding dairy cow. *Journal of Dairy Research*, 83(4): 438-441.
 18. Lisuzzo, A., Laghi, L., Fiore, F., Harvatine, K., Mazzotta, E., Faillace, V., Spissu, N., Zhu, C., Moscati, L., Fiore, E. 2022. Evaluation of the metabolomic profile through 1H-NMR spectroscopy in ewes affected by postpartum hyperketonemia. *Scientific Reports*, 12(1): 16463.
 19. Lisuzzo, A., Laghi, L., Faillace, V., Zhu, C., Contiero, B., Morgante, M., Mazzotta, E., Giancesella, M., Fiore, E. 2022. Differences in the serum metabolome profile of dairy cows according to the BHB concentration revealed by proton nuclear magnetic resonance spectroscopy (1H-NMR). *Scientific Reports*, 12(1):2525.
 20. Kersten, S., Desvergne, B., Wahli, W. 2000. Roles of PPARs in health and disease. *Nature*, 405:421-424.
 21. Yang, Y., Gocke, A.R., Lovett-Racke, A., Drew, P.D., Racke, M.K. 2008. PPAR Alpha Regulation of the Immune Response and Autoimmune Encephalomyelitis. *PPAR Res.*, 546753.
 22. Bobe, G., Young, J.W., Beitz, D.C. 2004. Invited review: pathology, etiology, prevention, and treatment of fatty liver in dairy cows. *J. Dairy Sci.*, 87:3105-3124.
 23. Wildeus, S. 2000. Current concept in synchronization of estrus: sheep and goats. *J. Anim. Sci.* 77:1-14.
 24. Sareminejad, P., Tabatabaei, S., Mamouei, M., Mirzadeh, K., Boujarpour, M. 2014. The effects of short and long term medroxyprogesterone acetate (MAP) sponge treatments on reproductive performance during the non-breeding season of Arabian ewes. *Iran J. Appl. Anim. Sci.*, 4:747-751.
 25. Özyurtlu, N., Ay, S.S., Küçükaslan, I., Güngör, O., Aslan, S. 2011. Effect of subsequent two short-term, short-term, and long-term progestagen treatments on fertility of Awassi ewes out of the breeding season. *Ankara Univ. Vet. Fak. Derg.*, 58:105-109.
 26. Yılmaz, Ç. 2015. The effects of melatonin and short-term progestagen treatments on reproductive performance in ewes out of the breeding season, Master Thesis, Ondokuz Mayıs University, Samsun, Turkey.
 27. Kutlu, M., Do an, H., Alkan, H., Serbest, U., Kutlu, H.R. 2022. Post-mating diclofenac vs. carprofen treatment on serum progesterone levels and reproductive outcomes in Hungarian-Merino ewes during the non-breeding season. *Reproduction in Domestic Animals*, 57:1529-1535.
 28. Rekawiecki, R., Kowalik, M.K., Slonina, D., Kotwica, J. 2008. Regulation of progesterone synthesis and action in bovine corpus luteum. *J. Physiol. Pharmacol.*, 59(9):75-89.
 29. Stocco, D.M., Clark, B.J. 1996. Regulation of the acute production of steroids in steroidogenic cells. *Endocr. Rev.*, 17:221-244.
 30. Niswender, G.D., Juengel, J.L., Silva, P.J., Rollyson, M.K., McIntush, E.W. 2000. Mechanisms controlling the function and life span of the corpus luteum. *Physiol. Rev.*, 80:1-29.
 31. Ashworth, C.J., Sales, D.L., Wilmut, I. 1984. Patterns of progesterone secretion and embryonic survival during repeated pregnancies in Damline ewes. Pages 741-743 in Proceedings of the 10th International Congress Animal Reproduction and Artificial Insemination, Urbana-Champaign, USA.
 32. Tamura, K., Ono, A., Miyagishima, T., Nagao, T., Urushidani, T. 2006. Profiling of gene expression in rat liver and rat primary cultured hepatocytes treated with peroxisome proliferators. *J. Toxicol. Sci.*, 31:471-490.
 33. Hotta, M., Nakata, R., Katsukawa, M., Hori, K., Takahashi, S., Inoue, H. 2010. Carvacrol, a component of thyme oil, activates PPAR and suppresses COX-2 expression. *J. Lipid Res.*, 51(1):132-139.
 34. Astakhova, A.A., Chistyakov, D.V., Pankevich, E.V., Sergeeva, M.G. 2015. Regulation of cyclooxygenase 2 expression by agonists of PPAR nuclear receptors in the model of endotoxin tolerance in astrocytes. *Biochemistry*, 80:1262-1270.
 35. Taniguchi, K., Kizuka, F., Tamura, I., Sugino, N. 2010. Prostaglandin F2-alpha Stimulates Cyclooxygenase-2 Expression and Prostaglandin F2-alpha Synthesis Through NF-kappaB Activation via Reactive Oxygen Species in the Corpus Luteum of Pseudopregnant Rats. *Biology of Reproduction*, 83:124-124.